

Adsorptive Pre-filtration to Increase Virus Filter Performance and Overall Process Robustness in Blood Derived Processes

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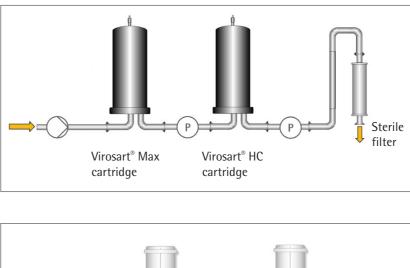
Adsorptive pre-filters

The evaluation of virus filters is not confined only to its capacity to retain viruses. Indeed, selection of a virus filter is influenced by numerous factors. One factor gaining increase importance is process economics. Different adsorptive pre-filters have been introduced to the marked for capacity increase of virus-retentive filters. Todays established adsorptive pre-filters are compared in the table below.

Depth Filter	CEX Membrane	Virosart [®] Max ¹
Nearly independent of conductivity	Affected by process conditions (pH, conductivity)	Performance independent from process conditions (conductivity)
High extractable	Low extractable	Low extractable
particle load	particle load	particle load
Integrity test	Integrity test	Integrity test by air
not available	not available	diffusion

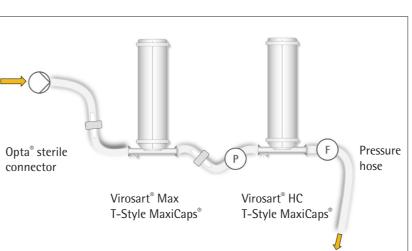
Implementation

Cartridges and capule format of the filter allows flexible process implementation:



Stainless steel housing setup

- Robust setup
- Steam sterilization and pre-use integrity testing possible



Single-use setup

- Ease of use
- Flexible
- Pre-use integrity testing limited under

¹ Sartorius patent DE102011105525-B4; US, EP and WO patents pending, 'Method for removing biopolymer aggregates and viruses from a fluid'

Characteristics of Virosart[®] Max



■ Virosart® HC 400 decay [L/m²] Virosart[®] MAX into Virosart[®] HC (1/1) 300 200 Capacity @ 100 5 g/l 10 g/l 20 g/l IVIG concentration

Working principle

- Combination of adsorptive capacity and size exclusion leads to removal of virus filter foulants
- Aggregates and | or small hydrophobic molecules are typical virus filter foulants

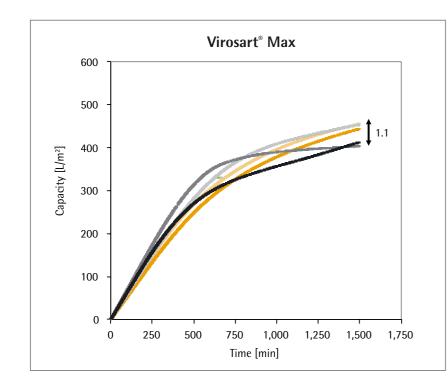
Filter Configuration

- Material: Optimized polyamide
- Pore size: 0.1 μm (nominal)
- Format: Triple-layer pleated elements
- Size: Available from 5 cm² to 30" elements

Higher capacity through aggregate reduction

The impact of Virosart[®] Max on the filtration of different IVIG concentrations (5, 10 and 20 g/L) through Virosart[®] HC 20 nm virus filter (5 cm² Minisart[®] devices) was analyzed. Filtrations have been performed with and without the use of Virosart[®] Max at 2.0 bar | 30 psi filtration pressure. Results were compared at 90% flow decay.

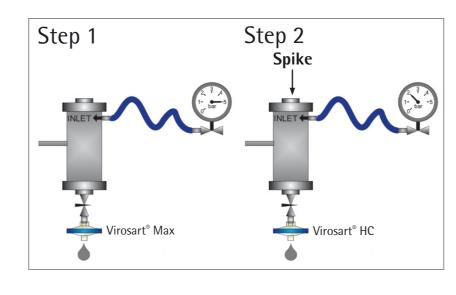
As a result, filtration capacity scales with solution concentration because the concentration of membrane fouling impurities scales accordingly.



Robust against process conditions The effect of different pre-filtration strategies was evaluated for IVIG (5 g/L) in different buffer conditions at varying pH and ionic strength using Virosart[®] HC 20 nm virus filter

Spiking studies

Spike





(decoupled)

- Product is pre-filtered off-line and afterwards virus spike is added to the product feed Pressure | flow adaption over pre-filter
- Low capacity of virus filter by highly fouling feed streams
- Common approach in the industry Pre-filtration before validation to restore
- sample

pre-filter.

Complex setup

viruses (MuLV, PRV)

Difficult control of feed titer

Alternative 3: Spiking virus selection

Accepted by regulatory authorities?

Validate virus-retentive filter for parvoviruses

(PPV, MVM) and imply sufficient LRV for larger

Alternative 1: In-line pre-filtration (coupled)

Pre-filter and virus filter are run in-inline and virus spike is added in-line.

- Virus retention by pre-filter not rated as robust
- Possible if pre-filter is tested independently for virus retention

Alternative 2: In-line pre-filtration with in-line spiking

Pre-filter and virus filter are run in-inline,

but the virus spike is added in-line after the

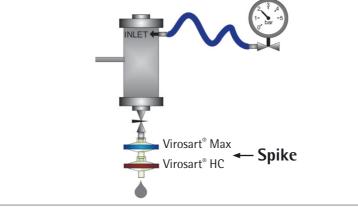
fully-contained sterile conditions

Automated setup

- Customized set-up
- High level of automation

(5 cm² Minisart[®] devices) at 2.0 bar | 30 psi.

As a result, the use of Virosart[®] Max results in lowest perfromacne spread by varying process conditions.



Virosart[®] Max

Virosart[®] HC

References

'Artifacts of Virus Filter Validation', P. Genest, H. Ruppach, C. Geyer, M. Asper, J. Parrella, B. Evans, A. Slocum, BioProcess International 2013.

